

REFERENCES

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Human cTnI Immunoassay

Catalog Number: SEKH-0400

For the quantitative determination of human cTnI concentrations
in cell culture supernates, serum, and plasma.

For research use only. Not for use in diagnostic procedures.

Country | Company: China | Beijing Solarbio Science & Technology Co.,Ltd
Address:NO.85A, Liandong U Valley, Tongzhou District, Beijing, P.R.China.
Tel: 86-10-56371241 Fax: 86-10-56371282 E-mail: service@solarbio.com

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RECOVERY: The coefficient of variation of both intra-assay and inter-assay were less than 10%.

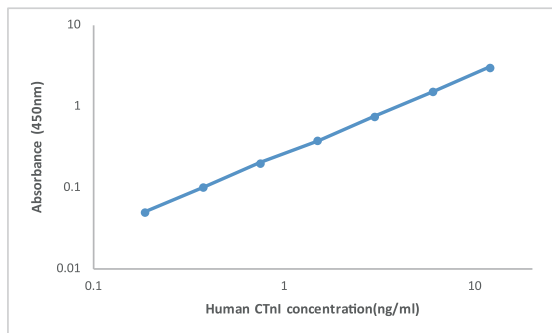
RECOVERY: The recovery of cTnI spiked to three different levels in four samples throughout the range of the assay in various matrices was evaluated.

Recovery of cTnI in two matrices

Sample Type	Average % of Expected Range(%)	Range(%)
Citrate plasma	90	85-95
Cell culture supernatants	102	96-108

LINEARITY: To assess the linearity of the assay, three samples were spiked with high concentrations of cTnI in various matrices and diluted with the appropriate Sample Diluent to produce samples with values within the dynamic range of the assay.

Dilution ratio	Recovery(%)	Citrate plasma	Cell culture supernatants
1:2	Average% of Expected	88	101
	Range(%)	83-93	97-105
1:4	Average% of Expected	95	110
	Range(%)	92-98	103-117



Performance Characteristics

SENSITIVITY: The minimum detectable dose was 0.9ng/mL.

SPECIFICITY: This assay recognizes both natural and recombinant human VCAM-1. The factors listed below were prepared at 50ng/ml in Standard /sample Diluent and assayed for cross-reactivity and no significant cross-reactivity or interference was observed.

Factors assayed for cross-reactivity

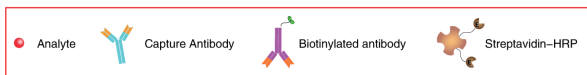
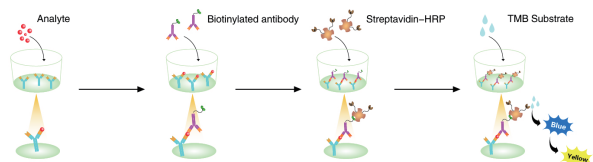
Recombinant human	Recombinant mouse	Recombinant porcine
EC-cTnI/PK1	PIGF-2	Rat cTnI 120(0.07%)
PDGF-AA	cTnI120	zebrafish cTnI 165
PDGF-AB	cTnI164	
PDGF-BB	cTnI-D	
PIGF	cTnI R2(KDR)/Fc Chimera	
cTnI-B167	cTnI R3(Flt-4) /Fc Chimera	
cTnI-C		
cTnI-D		
cTnI R3(Flt-4) /Fc Chimera		
EC-cTnI/PK1		

BACKGROUND

Troponin is a complex of three regulatory proteins (troponin C, troponin I, and troponin T) that is integral to muscle contraction in skeletal muscle and cardiac muscle, but not smooth muscle. Cardiac Troponin I (cTnI) is a protein that in felines is encoded by the TNNI3 gene. It is a tissue-specific subtype of troponin I. It has been shown to interact with PKD2L1, Troponin C type 1 and Polycystic kidney disease 2. An increased level of cTnI in the blood has been shown to be a biomarker of heart disorders, the most important of which is myocardial infarction (MI). Raised troponin levels indicate cardiac muscle cell death as the enzyme is released into the blood upon injury to the heart. It is measured in the blood to differentiate between unstable angina and MI (heart attack) in people with chest pain or acute coronary syndrome. cTnI can also indicate several forms of cardiomyopathy. In a community-based cohort study indicating the importance of silent cardiac damage, troponin I has been shown to predict mortality and first coronary heart disease event in men free from cardiovascular disease at baseline.

PRINCIPLE OF THE ASSAY

This assay employs the quantitative sandwich enzyme immunoassay technique. A monoclonal antibody specific for cTnI has been pre-coated onto a microplate. Standards and samples are pipetted into the wells and any cTnI present is captured by the coated antibody after incubation. Following extensive washing, a biotin-conjugate antibody specific for cTnI is added to detect the captured cTnI protein in sample. For signal development, horseradish peroxidase (HRP)-conjugated Streptavidin is added, followed by tetramethyl-benzidine (TMB) reagent. Following a wash to remove any unbound combination, and enzyme conjugate is added to the wells. Solution containing sulfuric acid is used to stop color development and the color intensity which is proportional to the quantity of bound protein is measurable at 450nm.



TECHNICAL HINTS AND LIMITATIONS

1. This Solarbio ELISA should not be used beyond the expiration data on the kit label.
2. To avoid cross-contamination, use a fresh reagent reservoir and pipette tips for each step.
3. To ensure accurate results, some details, such as technique, plasticware and water sources should be emphasized.
4. A thorough and consistent wash technique is essential for proper assay performance.
5. A standard curve should be generated for each set of samples assayed.
6. It is recommended that all standards and samples be assayed in duplicate.
7. Avoid microbial contamination of reagents and buffers. Buffers containing protein should be made under aseptic conditions and be prepared fresh daily.
8. In order to ensure the accuracy of the results, the standard curve should be made every time.

PRECAUTIONS

The Stop Solution suggested for use with this kit is an acid solution. Wear protective gloves, clothing, eye, and face protection. Wash hands thoroughly after handling.

CALCULATION OF RESULTS

1. The standard curve is used to determine the amount of specimens.
2. First, average the duplicate readings for each standard, control, and sample. All O.D. values are subtracted by the mean value of blank control before result interpretation.
3. Construct a standard curve by reducing the data using computer software capable of generating a four parameter logistic (4-PL) curve-fit. As an alternative, construct a standard curve by plotting the mean absorbance for each standard on the y-axis against the concentration on the x-axis and draw a best fit curve through the points on the graph.
4. The data may be linearized by plotting the log of the cTnI concentrations versus the log of the O.D. and the best fit line can be determined by regression analysis. This procedure will produce an adequate but less precise fit of the data. If samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.
5. This standard curve is provided for demonstration only. A standard curve should be generated for each set of samples assayed.

Typical data using the cTnI ELISA

Standardized(ng/ml)	OD.	OD.	Average	Corrected
0	0.082	0.09	0.086	---
1.875	0.178	0.167	0.172	0.086
3.75	0.221	0.225	0.223	0.137
7.5	0.336	0.352	0.344	0.258
1.5	0.525	0.546	0.535	0.449
3	0.954	0.934	0.944	0.858
6	1.362	1.339	1.350	1.264
12	2.159	2.182	2.170	2.084

ASSAY PROCEDURE

Prepare all reagents and standards as directed. Wash the plate 3 times before assay.



Add 100 μ l standard or samples to each well, shaking with Micro-oscillator (100r/min) to incubate 120 minutes at room temperature(25 \pm 2 $^{\circ}$ C).



Aspirate and wash 4 times

Add 100 μ l working solution of Biotin-Conjugate anti-human cTnI antibody to each well, shaking with Micro-oscillator (100r/min) to incubate 60 minutes at room temperature(25 \pm 2 $^{\circ}$ C).



Aspirate and wash 4 times

Add 100 μ l working solution of Streptavidin-HRP to each well, shaking with Micro-oscillator (100r/min) to incubate 30 minutes at room temperature (25 \pm 2 $^{\circ}$ C).



Aspirate and wash 5 times

Add 100 μ l Substrate solution to each well, incubate 5-30 minutes (depending on signal) at room temperature(25 \pm 2 $^{\circ}$ C). Protect from light.



Add 50 μ l Stop solution to each well. Read at 450nm within 5 minutes.

KIT COMPONENTS & STORAGE CONDITIONS

PART	SIZE	STORAGE OF OPENED/ RECONSTITUTED MATERIAL
Microwell Plate - antibody coated 96-well Microplate (8 wells \times 12 strips)	1 plate	Return unused wells to the foil pouch containing the desiccant pack. Reseal along entire edge of the zip-seal. May be stored for up to 1 month at 2 – 8 $^{\circ}$ C**
Standard - lyophilized, 12ng/ml upon reconstitution	2 vials	Aliquot and Store at -20 $^{\circ}$ C** for six months
Concentrated Biotin-Conjugated antibody(100X) - 120 μ l/vial	1 vial	Store at 2-8 $^{\circ}$ C ***for six months
Concentrated Streptavidin-HRP solution(100X) - 120 μ l/vial	1 vial	Store at 2-8 $^{\circ}$ C ***for six months
Standard /sample Diluent - 16 ml/vial	1 bottle	Store at 2-8 $^{\circ}$ C ***for six months
Biotin-Conjugate antibody Diluent - 16 ml/vial	1 bottle	Store at 2-8 $^{\circ}$ C ***for six months
Streptavidin-HRP Diluent - 16 ml/vial	1 bottle	Store at 2-8 $^{\circ}$ C ***for six months
Wash Buffer Concentrate (20x) - 30 ml/vial	1 bottle	Store at 2-8 $^{\circ}$ C ***for six months
Substrate Solution - 12 ml/vial	1 bottle	Store at 2-8 $^{\circ}$ C ***for six months
Stop Solution - 12 ml/vial	1 bottle	Store at 2-8 $^{\circ}$ C ***for six months
Plate Cover Seals	4 pieces	

**Provided this is within the expiration date of the kit.

OTHER SUPPLIES REQUIRED BUT NOT SUPPLIED

1. Microplate reader capable of measuring absorbance at 450 nm.
2. Pipettes and pipette tips.
3. Deionized or distilled water.
4. Squir bottle, manifold dispenser, or automated microplate washer.
5. 500 mL graduated cylinder.
6. Human cTnI controls (optional; available from Solarbio).

SPECIMEN COLLECTION & STORAGE

Cell Culture Supernates - Centrifuge cell culture media at 1000×g to remove debris. Assay immediately or aliquot and store samples at ≤ -20 °C. Avoid repeated freeze-thaw cycles.

Serum - Use a serum separator tube (SST) and allow samples to clot for 2 hours at room temperature or overnight at 2-8°C. Centrifuge at approximately 15 minutes at 1000×g. Assay immediately or aliquot and store samples at ≤ -20 °C. Avoid repeated freeze-thaw cycles.

Plasma - Collect plasma using EDTA, heparin, or citrate as an anticoagulant. Centrifuge for 15 minutes at 1000×g within 30 minutes of collection. Assay immediately or aliquot and store samples at ≤ -20 °C. Avoid repeated freeze-thaw cycles.

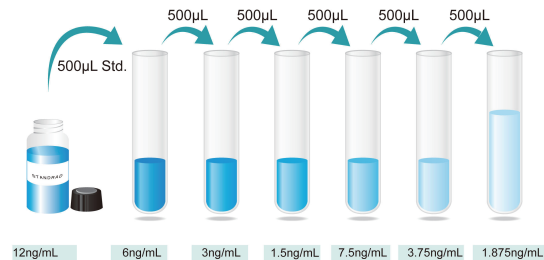
Note: It is recommended to conduct a pre-test before the formal experiment to determine the dilution ratio.

REAGENTS PREPARATION

1. **Temperature returning** - Bring all kit components and specimen to room temperature (20-25°C) before use.
2. **Wash Buffer** - Dilute 30mL of Wash Buffer Concentrate with 570mL of deionized or distilled water to prepare 600mL of Wash Buffer. If crystals have formed in the concentrate Wash Buffer, warm to room temperature and mix gently until the crystals have completely dissolved.

3. **Standard/Specimen** - Reconstitute the Standard with 1mL of Standard/Sample Diluent. This reconstitution produces a stock solution of 12ng/mL. Allow the standard to sit for a minimum of 15 minutes with gentle agitation prior to making dilutions. Pipette 500 μ L of Standard/Sample Diluent into 6ng/ml tube and the remaining tubes. Use the stock solution of 12ng/mL to produce a 2-fold dilution series (below). Mix each tube thoroughly and change pipette tips between each transfer. The 12ng/mL standard serves as the high standard. The Standard/Sample Diluent serves as the zero standard (0 pg/mL).

***If you do not run out of re-melting standard, store it at -20°C. Diluted standard shall not be reused.**



Preparation of cTnI standard dilutions

4. **Working solution of Biotin-Conjugate anti-human cTnI antibody:** Make a 1:100 dilution of the concentrated Biotin-Conjugate solution with the Biotin-Conjugate antibody Diluent in a clean plastic tube.
***The working solution should be used within one day after dilution.**
5. **Working solution of Streptavidin-HRP:** Make a 1:100 dilution of the concentrated Streptavidin-HRP solution with the Streptavidin-HRP Diluent in a clean plastic tube.
***The working solution should be used within one day after dilution.**