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Canine IgG Immunoassay

Catalog Number: SEKC-0050

For the quantitative determination of Canine IgG concentrations in cell culture supernates, serum, and plasma.

For research use only. Not for use in diagnostic procedures.

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REPEATABILITY: The coefficient of variation of both intra-assay and inter-assay were less than 10%.

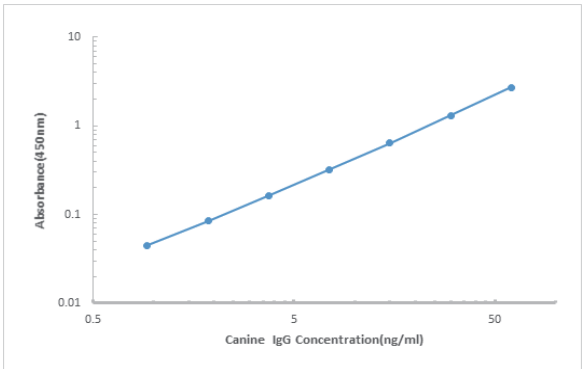
RECOVERY: The recovery of IgG spiked to three different levels in four samples throughout the range of the assay in various matrices was evaluated.

Recovery of IgG in two matrices

Sample Type	Average % of Expected Range (%)	Range (%)
Citrate plasma	91	83-101
Cell culture supernatants	93	85-103

LINEARITY: To assess the linearity of the assay, three samples were spiked with high concentrations of IgG in various matrices and diluted with the appropriate Sample Diluent to produce samples with values within the dynamic range of the assay. (The plasma samples were initially diluted 1:1)

Dilution ratio	Recovery(%)	Citrate plasma	Cell culture supernatants
1:2	Average% of Expected	95	104
	Range (%)	87-102	96-115
1:4	Average% of Expected	96	103
	Range (%))	89-107	96-113



Representative standard curve for IgG ELISA.

Performance Characteristics

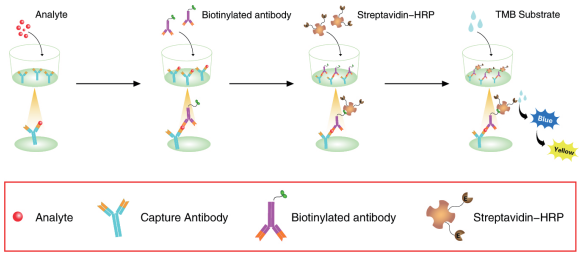
SENSITIVITY: The minimum detectable dose was 900pg/mL.
SPECIFICITY: This assay recognizes both natural and recombinant Canine IgG. The factors listed below were prepared at 10ng/ml in Standard /sample Diluent and assayed for cross-reactivity and no significant cross-reactivity or interference was observed.
BMP1, BMP2, BMP4, IL-1β, IL-1ra, IL-2, IL-4, IL-5, IL-6, IL-8, IL-10, IL-12, IL-13, IL-15,
IFN-γ, TGFβ1, TGFβ2, TGFβ3, TLR1, TLR2, TLR3, TNF-α, VEGF.

BACKGROUND

Immunoglobulin G (IgG) is antibody molecule. Each IgG is composed of four peptide chains — two heavy chains γ and two light chains. IgG antibodies are large molecules of about150 kDa composed of four peptide chains. It contains two identical class γ heavy chains of about 50 kDa and two identical light chains of about 25 kDa, thus a tetrameric quaternary structure.The two heavy chains are linked to each other and to a light chain each by disulfide bonds. The resulting tetramer has two identical halves, which together form the Y-like shape. Each end of the fork contains an identical antigen binding site. The Fc regions of IgGs bear a highly conserved N-glycosylation site. The N-glycans attached to this site are predominantly core fucosylated diantennary structures of the complex type. In addition, small amounts of these N glycans also bear bisecting GlcNAc and α-2, 6-linked sialic acid residues . IgG antibodies are involved in predominantly the secondary immune response.

PRINCIPLE OF THE ASSAY

This assay employs the quantitative sandwich enzyme immunoassay technique. A monoclonal antibody specific for IgG has been pre-coated onto a microplate. Standards and samples are pipetted into the wells and any IgG present is captured by the coated antibody after incubation. Following extensive washing, a biotin-conjugate antibody specific for IgG is added to detect the captured IgG protein in sample. For signal development, horseradish peroxidase (HRP)-conjugated Streptavidin is added, followed by tetramethyl-benzidine (TMB) reagent. Following a wash to remove any unbound combination, and enzyme conjugate is added to the wells. Solution containing sulfuric acid is used to stop color development and the color intensity which is proportional to the quantity of bound protein is measurable at 450nm.



TECHNICAL HINTS AND LIMITATIONS

1.This Solarbio ELISA should not be used beyond the expiration data on the kit label.
2.To avoid cross-contamination, use a fresh reagent reservoir and pipette tips for each step.
3.To ensure accurate results, some details, such as technique, plasticware and water sources should be emphasized.
4.A thorough and consistent wash technique is essential for proper assay performance.
5.A standard curve should be generated for each set of samples assayed.
6.It is recommended that all standards and samples be assayed in duplicate.
7.Avoid microbial contamination of reagents and buffers. Buffers containing protein should be made under aseptic conditions and be prepared fresh daily.
8.In order to ensure the accuracy of the results, the standard curve should be made every time.

PRECAUTIONS

The Stop Solution suggested for use with this kit is an acid solution. Wear protective gloves, clothing, eye, and face protection. Wash hands thoroughly after handling.

CALCULATION OF RESULTS

1.The standard curve is used to determine the amount of specimens.
2.First, average the duplicate readings for each standard, control, and sample. All O.D. values are subtracted by the mean value of blank control before result interpretation.
3.Construct a standard curve by reducing the data using computer software capable of generating a four parameter logistic (4-PL) curve-fit. As an alternative, construct a standard curve by plotting the mean absorbance for each standard on the y-axis against the concentration on the x-axis and draw a best fit curve through the points on the graph.
4.The data may be linearized by plotting the log of the IgG concentrations versus the log of the O.D. and the best fit line can be determined by regression analysis. This procedure will produce an adequate but less precise fit of the data. If samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.
5.This standard curve is provided for demonstration only. A standard curve should be generated for each set of samples assayed.

Typical data using the IgG ELISA

Standared(ng/ml)	OD.	OD.	Average	Corrected
0	0.045	0.048	0.046	----
1.875	0.094	0.096	0.095	0.048
3.75	0.187	0.193	0.190	0.143
7.5	0.315	0.331	0.323	0.276
15	0.525	0.541	0.533	0.486
30	0.866	0.847	0.856	0.810
60	1.548	1.562	1.555	1.508
120	2.768	2.745	2.756	2.710

***The working solution should be used within one day after dilution.**

5. **Working solution of Streptavidin-HRP(120μL) -**Centrifuge for 1 min at 6000 x g to bring down the material prior to open the vial. The vial contains 120 μL HRP Conjugate sufficient for a 96-well plate.Make 1:100 dilutions in Reagent Diluent. If the entire 96-well plate is used, add 100 ul of HRP Conjugate to 10 mL of Streptavidin-HRP Diluent to make working dilution of HRP Conjugate and mix thoroughly prior to the assay. The rest of undiluted HRP Conjugate can be stored at 4°C for up to 6 months. DO NOT FREEZE.

***The working solution should be used within one day after dilution.**

ASSAY PROCEDURE

Prepare all reagents and standards as directed. Wash the plate 3 times before assay.

⇩

Add 100μl standard or samples to each well, shaking with Micro-oscillator (100r/min) to incubate 60 minutes at room temperature(25±2℃).

⇩ Aspirate and wash 4 times

Add 100μl working solution of Biotin-Conjugate anti-Canine IgG antibody to each well, shaking with Micro-oscillator (100r/min) to incubate 60 minutes at room temperature (25±2℃).

⇩ Aspirate and wash 4 times

Add 100μl working solution of Streptavidin-HRP to each well, shaking with Micro-oscillator (100r/min) to incubate 20 minutes at room temperature(25±2℃).

⇩ Aspirate and wash 5 times

Add 100μl Substrate solution to each well, incubate 5-20 minutes (depending on signal) at room temperature(25±2℃).Protect from light.

⇩

Add 50μl Stop solution to each well. Read at 450nm within 5 minutes.

KIT COMPONENTS& STORAGE CONDITIONS

PART	SIZE	STORAGE OF OPENED/ RECONSTITUTED MATERIAL
Microwell Plate - antibody coated 96-well Microplate (8 wells ×12 strips)	1 plate	Return unused wells to the foil pouch containing the desiccant pack. Reseal along entire edge of the zip-seal. May be stored for up to 1 month at 2 – 8℃**
Standard-lyophilized,120ng/ml upon reconstitution	2 vials	Aliquot and Store at -20°C** for six months
lyophilized Biotin-Conjugated antibody	1 vial	Store at 2-8°C **for six months
Concentrated Streptavi-din-HRP	1 vial	Store at 2-8°C **for six months
Standard /sample Diluent	1 bottle	Store at 2-8°C **for six months
Biotin-Conjugate antibody Diluent	1 bottle	Store at 2-8°C **for six months
Streptavidin-HRP Diluent	1 bottle	Store at 2-8°C **for six months
20 x Wash Buffer Concentrate	1 bottle	Store at 2-8°C **for six months
Substrate Solution	1 bottle	Store at 2-8°C **for six months
Stop Solution	1 bottle	Store at 2-8°C **for six months
Plate Cover Seals	4 pieces	

**Provided this is within the expiration date of the kit.

OTHER SUPPLIES REQUIRED BUT NOT SUPPLIED

1.Microplate reader capable of measuring absorbance at 450 nm.
2.Pipettes and pipette tips.
3.Deionized or distilled water.
4.Squirt bottle, manifold dispenser, or automated microplate washer.
5.500 mL graduated cylinder.

SPECIMEN COLLECTION & STORAGE

Cell Culture Supernates - Centrifuge cell culture media at 1000×g to remove debris. Assay immediately or aliquot and store samples at ≤ -20 °C. Avoid repeated freeze-thaw cycles.

Serum - Use a serum separator tube (SST) and allow samples to clot for 2 hours at room temperature or overnight at 2-8℃. Centrifuge at approximately for 15 minutes at 1000×g. Assay immediately or aliquot and store samples at ≤ -20 °C. Avoid repeated freeze-thaw cycles.

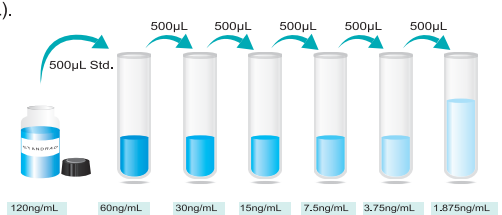
Plasma - Collect plasma using EDTA, heparin, or citrate as an anticoagulant. Centrifuge for 15 minutes at 1000×g within 30 minutes of collection. Assay immediately or aliquot and store samples at ≤ -20 °C. Avoid repeated freeze-thaw cycles.

Note: It is recommended to conduct a pre-test before the formal experiment to determine the dilution ratio.

REAGENTS PREPARATION

1. **Temperature returning** - Bring all kit components and specimen to room temperature (20-25℃) before use.
2. **Wash Buffer** - Dilute 30mL of 20x Wash Buffer Concentrate with 570mL of deionized or distilled water to prepare 600mL of Wash Buffer. If crystals have formed in the concentrate Wash Buffer, warm to room temperature and mix gently until the crystals have completely dissolved.
3. **Standard\Sample (2 vials)** - Canine IgG Standard has a total of 2 vials. Each vial contains the standard sufficient for generating a standard curve. Reconstitute the Standard with 1.0mL of Standard/Sample

Diluent. This reconstitution produces a stock solution of 120 ng/mL. Allow the standard to sit for a minimum of 15 minutes with gentle agitation prior to making dilutions.Pipette 500µL of Standard/Sample Diluent into 60ng/ml tube and the remaining tubes. Use the stock solution of 120ng/mL to produce a 2-fold dilution series (below). Mix each tube thoroughly(vortex 20 sec for each of dilution step) and change pipette tips between each transfer. The 120 ng/mL standard serves as the high standard. The Standard/sample Diluent serves as the zero standard (0 ng/mL).



Preparation of Canine IgG standard dilutions

***If you do not run out of re-melting standard, store it at -20℃. Diluted standard shall not be reused.**

4. **Working solution of Biotin-Conjugate anti-Canine IgG antibody (1vials)-**The lyophilized Detection Antibody should be stored at 4℃ to -20℃ in a manual defrost freezer for up to 6 months, if not used immediately. Centrifuge for 1 min at 6000 x g to bring down the material prior to open the vial. The vial contains sufficient Detection Antibody for a 96-well plate. Add 110 µL of sterile Bioutin-Conjugate antibody Diluent to each vial and vortex 30 sec to obtain the stock solution. If the entire 96-well plate is used, take 25µL of detection antibody stock solution into 10 mL of Biotin-Conjugate antibody Diluent to make working dilution of Detection Antibody and mix thoroughly prior to the assay. If the partial antibody is used. make a 1:400 dilution of the concentrated Biotin-Conjugate solution with the Biotin-Conjugate antibody Diluent in a clean plastic tube.